

# New Approaches for Estimating Risk from Exposure to Diethylstilbestrol

Gerald R. Cunha,<sup>1</sup> John-Gunnar Forsberg,<sup>2</sup> Robert Golden,<sup>3</sup> Arthur Haney,<sup>4</sup> Taisen Iguchi,<sup>5</sup> Retha Newbold,<sup>6</sup> Shanna Swan,<sup>7</sup> and Wade Welshons

<sup>1</sup>Anatomy Department and Reproductive Endocrinology Center, University of California, San Francisco, California USA; <sup>2</sup>Tornblad Institute Lund, Sweden; <sup>3</sup>ToxLogic, Potomac, Maryland USA; <sup>4</sup>Department of Obstetrics and Gynecology, Duke University Medical Center, Durham, North Carolina USA; <sup>5</sup>Department of Biology and Graduate School of Integrated Science, Yokohama City University, Yokohama, Japan; <sup>6</sup>Laboratory of Toxicology, Environmental Toxicology Program, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina USA; <sup>7</sup>Department of Family and Community Medicine, University of Missouri, Columbia, Missouri USA; <sup>8</sup>Department of Veterinary Biomedical Science, University of Missouri, Columbia, Missouri USA

A subgroup from a National Institute of Environmental Health Sciences, workshop concerned with characterizing the effects of endocrine disruptors on human health at environmental exposure levels considered the question, If diethylstilbestrol (DES) were introduced into the market for human use today and likely to result in low-dose exposure of the human fetus, what would be required to assess risk? On the basis of an analysis of the quality of data on human DES exposure, the critical times and doses for inducing genital tract malformations and cancer must be determined. This would be facilitated through analysis of the ontogeny of estrogen receptor expression in the developing human genital tract. Models of low-dose estrogenic effects will have to be developed for human and rodent genital tract development. Mouse models offer many advantages over other potential animal models because of the wealth of the earlier literature, the availability of sensitive end points, the availability of mutant lines, and the possibility of generating genetically engineered model systems. Through multidisciplinary approaches, it should be possible to elucidate the cellular and molecular mechanisms of endocrine disruption elicited by estrogens during development and facilitate an assessment of risk to humans. *Key words:* carcinogenesis, clear cell carcinoma, diethylstilbestrol (DES), genital tract, human, teratogenesis. — *Environ Health Perspect* 107(suppl 4):625–630 (1999).

<http://ehpnet1.niehs.nih.gov/docs/1999/suppl-4/625-630cunha/abstract.html>

This report is the product of a subgroup from a National Institute of Environmental Health Sciences, workshop concerned with characterizing the effects of endocrine disruptors on human health at environmental exposure levels. This workshop provided a forum to discuss methods and data needed to improve risk assessments of endocrine disruptors. This report addresses data on the health effects of diethylstilbestrol (DES) and how this information may be used to evaluate risks from exposure to weaker synthetic estrogens. The goal of this review is a re-evaluation of the risk assessment of the human DES experience, using the abundant experimental animal data to answer the following questions: How can we use the human and animal data to better anticipate adverse health effects from agents that are introduced in the future? How could we have anticipated the consequences of DES exposure from the information available when DES was approved for use in pregnant women? Can general lessons be drawn regarding animal-to-human extrapolation for endocrine disruptors? To answer these questions, a historical perspective is required.

Years before clinical use of DES in pregnant women, estrogens in general, and DES specifically, were known to induce breast cancer in postnatal mice and rats when pharmacologic doses were given chronically over long periods (1–3). The relationship of postnatal studies to possible transplacental

carcinogenesis was certainly not appreciated in the 1930s and early 1940s. Indeed, transplacental carcinogenesis of DES or other estrogens was not considered or reported by investigators at that time. Although prior to 1945, estrogens were known to perturb urogenital development in fetal rodents (4,5) and were thought to cross the placenta in humans (6,7), direct evidence of teratogenicity of estrogens in humans was unknown until after the association between DES and vaginal adenocarcinoma was reported. In any case, despite the tragedy of the DES episode, the human DES clinical data offer an unprecedented opportunity to learn about the consequences of *in utero* exposure to a potent estrogen and thus to infer potential risks following exposure to less potent environmental estrogens.

If properly interpreted, lessons from the DES episode may prove invaluable for judging potential effects of compounds that have been or will be identified as potential endocrine disruptors. It will be important, however, to keep in mind the considerable differences in potency between such compounds when inferences are drawn concerning potential effects. For example, the carcinogenicity of DES was identified in a human study including only 8 cases and 32 controls (8). Normal sample size calculations would rule such a study as inadequate. However, because the cancer induced by

DES (clear cell vaginal adenocarcinoma) was so rare in young women, the association between prenatal DES exposure and development of clear cell adenocarcinoma of the vagina was easily identified. Clearly, most chemicals with significantly less potent endocrine (e.g., estrogenic) effects will convey much-reduced risks, particularly at low doses. Thus, study designs for other endocrine disruptors will have to be more precise and more powerful, especially if the background incidence of a particular lesion is substantial. Given the history of the DES episode, we have considered this issue: If DES were introduced into the market for human use today and were likely to result in low-dose exposure of the human fetus, what would be required to assess risk of developing adverse health outcomes such as cancer or impaired reproductive potential? To answer this question, we have considered the following points:

- *Delineation of the critical times and doses for inducing genital tract malformations and cancer.* Critical time periods and doses must be determined for DES-induced malformations of the developing human Mullerian duct, Wolffian duct, urogenital sinus, and their organ derivatives. Given the plethora of teratogenic effects of DES on the developing male and female genital tracts (Tables 1,2), different lesions are expected to have different periods of temporal susceptibility as well as different dose levels required to induce such effects. The background information for such teratogenic studies can be estimated from the literature of classical embryologic studies. Retrospective studies on the incidence of

This report was developed at the Workshop on Characterizing the Effects of Endocrine Disruptors on Human Health at Environmental Exposure Levels held 11–13 May 1998 in Raleigh, North Carolina.

Address correspondence to G.R. Cunha, Anatomy Department and Reproductive Endocrinology Center, University of California San Francisco, CA 94143. Telephone: (415) 476-4140. Fax: (415) 502-2270. [grcunha@itsa.ucsf.edu](mailto:grcunha@itsa.ucsf.edu)

This work was funded in part by the following NIH grants: DK02397, DK51101, DK45861, DK52708, CA64872, CA59831, DK52721, DK51397, AG13784, AG15500.

Received 25 September 1998; accepted 24 March 1999.

**Table 1.** Effects of perinatal exposure to estrogenic substances in males.

Rat	Mouse	Human	Estrogenic effects	Notes
+	+	Not known	Decreased prostate and seminal vesicle secretion	High doses
+	+	Not known	Increased prostate weight	Low doses
Not known	+/-	+/-	Rete testes tumor	
Not known	+	+/-	Decreased sperm count	
Not known	+	Not known	Increased lactoferrin in seminal vesicle	High doses
+	+	+	Mullerian duct remnants in males, hypertrophic prostatic utricle	
Not known	+	+/-	Impaired reproduction	High doses
Not known	+	+	Anomalies of male external genitalia	
Not known	+	Not known	Thyroid effects	
Not known	+	Not known	Intersex	
Not known	+	Not known	Alterations in sex behavior	
+	+	-	Hypothalamic changes	

**Table 2.** Effects of perinatal exposure to estrogenic substances in females.

Rat	Mouse	Human	Estrogenic effects	Notes
+	+	Not known	Polyovulatory follicles	
+	+	Not relevant	Ovary-independent vaginal cornification	
-	+	+	Cervical/vaginal carcinoma	
Not known	+	+/-	Uterine carcinoma	
Not known	+	+/-	Immune dysfunction	More frequent autoimmune disease in women
+	+	Not known	Polycystic ovary	
+	+	+/-	Estrus/menstrual cycle disturbance	
+	+	Not known	Mammary tumors in daughters	Small increased risk in mothers
+	+	+	Vaginal adenosis	
+	+	+	Impaired myometrial development	T-shaped uterus in humans
Not known	+	+	Impaired reproduction	
+	+	Not known	Mammary gland alterations	
+	+	+	Mesonephric remnants	
Not known	+	Not known	Accelerated vaginal opening	Equivalent to puberty in humans
Not known	+	+	Oviductal malformations	
Not known	+	+	Uterine and cervical malformations	

certain lesions such as adenosis in women exposed *in utero* to DES in relationship to the initiation of DES treatment are helpful in establishing periods of susceptibility even though estimates of gestational ages of exposure may be inexact. Human cell lines are not particularly useful for determining the critical times and doses for inducing human genital tract malformations. However, analysis of DES effects on grafts of human fetal genital tracts in nude mice has proved to be a reliable method for assessing periods of susceptibility for the induction of different lesions (9–13). Indeed, this method is perhaps the only method for delineating the critical times and doses for inducing human genital tract malformations. Contrary to popular thought, a considerable amount of abortus material is available for such screening purposes. Clearly, it will be necessary to extend these studies to determine the specific times and doses of DES required to induce various lesions in the developing human genital tract. This will provide a relevant basis for judging the likelihood of similar effects from compounds identified as environmental endocrine disruptors. Recognizing that other developing organ

systems such as the neuroendocrine and immune systems are also sensitive to estrogenic substances, critical periods for inducing adverse effects on these systems would also need to be established.

- *Ontogeny of estrogen receptor (ER) expression in the developing human genital tract.* The presumed mechanism of action of DES is through ER in the developing genital tract, even though action via other receptors and nonreceptor-mediated mechanisms should also be considered. Although the ontogeny of expression of ER has been studied in the mouse and rat, data are meager for the human genital tract (14–16). Future studies need to take into account both ER-alpha (ER $\alpha$ ) and the recently discovered ER-beta (ER $\beta$ ) (17). The literature is even more deficient for the ontogeny of androgen receptors and progesterone receptors, especially in the human fetal genital tract (18,19). Such ontogenetic studies will be critical for the interpretation of adverse effects of DES and other chemicals and drugs with hormonal activity.
- *Low-dose models of estrogenic effects on genital tract development.* Even though DES exposure levels of the developing

human vary considerably, all clinical exposure of human fetuses falls into the high-dose range. High-dose animal studies mainly designed to duplicate the therapeutic doses prescribed for pregnant women have been extensively published for mouse and rat, although a few low-dose studies have been described (20–22). Low-dose animal models are now receiving attention (23–25) and indicate that outcomes from high- and low-dose exposure can be both qualitatively and quantitatively different. Additional low dose–response animal work is required to assess the potential effects of less potent estrogenic compounds on the many end points previously described (Tables 1,2). Because of the wealth of DES data in both animals and humans, an animal model that is sensitive to DES effects at low doses may be useful in screening other compounds with potentially similar mechanisms of action. The ability to study low-dose effects on human urogenital tract development would be valuable. Such information in the human could be obtained by studying transplants of human fetal genital tracts in DES-treated nude mouse hosts (10). The transplant model of human fetal genital tracts could be used as either a screen or a confirmatory test for low-dose effects seen in animal studies. Such proposed use of human fetal genital tracts in DES-treated nude mouse hosts to delineate the lower end of the dose response in developing human genital tracts could provide the basis for assessing low-dose effects of environmental chemicals identified as having potential endocrine-disrupting effects.

- *Characterization of human fetal serum-binding proteins.* Calculation of relative binding affinity (RBA) and serum-modified access (SMA) is a powerful method for determining levels of free compound capable of eliciting estrogenic effects in a test system (26). This information is available for the fetal rat and mouse but is not available for the human fetus. Umbilical cord serum from full-term fetuses should be analyzed to assess RBA and SMA. Serum from first and second trimester human fetuses will be extremely difficult to obtain. Fetal primate serum may be a useful substitute.

### What Is the Quality of Human DES Exposure Data?

Even though the dosing regimen recommended by Smith et al. (27) was in widespread use, this dosage pattern was far from universal, particularly since its efficacy had never been established. Thus, there are defined cohorts of women who received 1.4–17.9 g DES as a total dose during

pregnancy (28), even though a certain level of imprecision exists concerning timing of exposure and the numbers of individuals exposed *in utero* to specific maternal doses of DES. In any case, the human data available are primarily related to the high-dose regimens. With respect to clear cell vaginal adenocarcinoma, it is evident that for many patients even these high-dose exposures were insufficient to induce neoplasia in all but a small subset of patients. Thus, it would appear that there are dosages and/or periods of high-dose DES exposure that do not trigger neoplastic change in both humans and animals. For nonmalignant lesions in humans such as adenosis, cervical defects, and T-shaped uteri, the timing of exposure is important in generating genital tract abnormalities. Based upon the abundant human clinical data, the relationship between dosage and the development of nonmalignant lesions suggests that there are also DES doses below which adverse noncancer effects are not seen. However, for humans especially, there is a great need to accurately define the exact dose range and timing that elicit genital tract malformations and those doses that are below the threshold for eliciting adverse effects. Use of a nude mouse transplant system for human fetal genital tracts may be the only method to obtain this critical data. Acquisition of these types of data will permit relevant potency comparisons between DES and environmental compounds identified as having estrogenic activity.

The existence of cohorts exposed to DES at mean total maternal doses spanning more than an order of magnitude provide an opportunity to study the dose-response characteristics for relatively high-dose DES-induced effects. These data also provide the opportunity to compare human clinical data and dose-response data for DES-induced effects observed in animal studies. As part of the ongoing follow-up of DES-exposed cohorts, substantial numbers of exposed males and females have been studied. Depending on the timing of exposure and the total maternal DES dose administered, unequivocal (and more readily observed) effects seen in males and females include reproductive tract malformations, impaired reproduction, and vaginal carcinoma (29–33). More equivocal effects include decreased sperm count (34–36), immune dysfunction (37), alterations in sexual behavior (38), disturbed menstrual cycles (39), and testicular cancer (40). To date, DES-exposed males and females diagnosed with malignant or nonmalignant lesions include individuals in childhood and puberty, and adults less than 50 years of age. Substantial numbers of males and females in the DES adenosis cohort are just now reaching 50 years of age. This is the age when male and female reproductive tract neoplasias typically begin to

occur. Additional follow-up of DES-exposed sons will be essential to establish whether they are at increased risk of testicular or prostate cancer. It is important to note that, in general, women have been more extensively studied than men because of the initial association between *in utero* DES exposure and vaginal cancer and also because of a greater interaction of women with health care services and providers. Furthermore, women have formed DES support groups and have successfully lobbied the government for studies of the adverse effects of DES.

It should also be noted that some of the adverse effects observed as a consequence of *in utero* exposure to DES occur against an extremely low background incidence of reproductive tract malformations and vaginal carcinoma. By following the DES-exposed cohorts, it will be challenging to determine if more prevalent conditions (i.e., thyroid effects, breast and prostate cancer, endometriosis, immune dysfunction) or conditions that increase in frequency with age (e.g., declining immune function, endometrial hyperplasia and cancer, ovarian cancer, benign prostatic hyperplasia, prostatic cancer) are increased as a result of *in utero* exposure to DES. Also, the possibility of third-generation effects has to be considered (41).

The lack of low-dose human DES exposure data might be addressed by the use of human fetal reproductive tract tissue transplanted to nude mouse hosts. This would permit a detailed study of the lower end of the dose-response curve. Previous use of such a model system of human fetal reproductive tract transplants has demonstrated that many of the high-dose DES effects observed in the epidemiologic studies can be induced experimentally in such transplants of human fetal reproductive tracts. The transplant model offers the possibility of extending dose-response studies in the human well below the DES doses used clinically. Additionally, such data would also serve as a bridge between the low-dose mouse data and potential low-dose human effects data. Because of the known potency of DES, acquiring these kinds of data would provide the most relevant basis for judging whether compounds identified as having estrogenic activity might be expected to be teratogenic in humans.

### Possible Pharmaceutical Exposures to Endocrine-Disrupting Chemicals during Pregnancy

Environmental contamination by endocrine-disrupting agents has received considerable attention in the scientific and lay press, and the impacts of such agents on reproduction in wildlife has had a deleterious impact on many

species (42). Humans can be exposed to endocrine disruptors through use of a variety of commonly used medications. Thus, potentials for exposing women of reproductive age to hormonally active drugs (estrogens, androgens, or progestins) include the following possibilities: *a*) inadvertent use of a drug in the luteal phase of a conceptive cycle, *b*) inadvertent administration of a drug during pregnancy in oligo/amenorrheic women, *c*) contraceptive failure coupled with continued use of birth control pills, *d*) inadvertent administration of a drug following non-hormonal contraceptive failure (intrauterine devices, condoms, diaphragms), and *e*) use of a drug in gynecologic or medical disease in women of child-bearing age. Inappropriate exposure to estrogens, androgens, and/or progestins can elicit severe malformations of the genital tract. Thus, low- or high-dose exposure to hormonally active compounds should be avoided at all costs, and if exposure occurs, any adverse outcomes should be monitored. Some currently used pharmaceuticals that may pose risks to the human fetus are given in Table 3.

## Animal Models

### Summary of the Models

The animal models in which the developmental effects of DES exposure have been studied mainly include the mouse, rat, and hamster. The mouse has been the most extensively studied species, and the size of the data set in mice is superior to those for all other species combined. This extensive body of evidence in the mouse extends back about 50 years. The perinatal DES-treated mouse model correlates remarkably well with the adverse effects observed in both male and female humans exposed *in utero* to DES. Tables 1 and 2 illustrate these effects and the correspondence in effects between rodent and human studies.

### The Mouse As the Best Animal Model

Although the rat and hamster may be equally appropriate for modeling DES effects in humans, data in these species are not nearly as abundant as those for the mouse. Rat studies are only likely to further validate the existing mouse data. Another advantage of the mouse model is the wealth of genetic information that is available and the relative ease of using transgenic and gene knockout mice to study the mechanism by which DES produces adverse effects. In addition, genetically modified strains of mice might make it possible to study the interaction between direct DES effects, immune factors, and endogenous hormones in teratogenic or carcinogenic processes. Additional advantages of the mouse model include the

**Table 3.** Drugs potentially capable of eliciting adverse effects on development via disturbance of sex hormones.

Combination estrogen/progestin (oral contraceptives and continuous combined hormone replacement therapy, continuous combined oral contraceptives)
C-19 norprogestins (modified testosterone)
High androgenicity - levonorgestrel
Intermediate androgenicity - norethindrone (acetate), ethynodiol diacetate, norgestrol
Low androgenicity - gestidine, norgestimate, desogestrel
C-21 progestins: medroxyprogesterone acetate
Estrogens
Ethinyl estradiol (20–35 µg)
Mestranol (35 µg)
Conjugated estrogens (0.625–2.5 mg)
Progestin-only products
Medroxyprogesterone acetate (oral and/or depo)
Megestrol acetate (Megace) - for endometrial cancer
Norethindrone acetate (Micronor)
Levonorgestrel implants (Norplant)
Micronized progesterone (oral)
Progesterone transvaginal gel
Progesterone in oil for injection
Antiandrogens
Spironolactone (receptor antagonist and synthesis inhibitor)
Flutamide (receptor antagonist)
Finasteride (5 $\alpha$ -reductase inhibitor)
Cimetidine (tagamet, significant effects within the range of ulcer treatment)
Cyproterone acetate (formulated with ethanyl estradiol in Europe)
Letozole (aromatase inhibitor)
Androgens
Danazol (androgen/anabolic for treatment of endometriosis, hereditary angioneurotic edema, fibrocystic breast disease, AIDS)
Estratest (1.25 mg + 2.5 mg methyltestosterone)
Depotestadiol (estradiol + testosterone for intramuscular use)
Testosterone (implants/gel/transdermal)
Oxandrolone (and other anabolic agents)
Estrogens
Conjugated estrogens
Conjugated equine estrogens (equillin, equillin, other equine estrogens)
Micronized estradiol
Transdermal estradiol
Transvaginal estradiol
SERMs
DES
Clomiphene citrate
Tamoxifen
Nafoxidine
Raloxifene
Toremifene
Many others in development
Antiprogestins
RU486 (receptor blocker)
Epostane (synthesis inhibitor)
Other agents to which fetuses could be inadvertently exposed
Fluconazole (systemic antifungal)
Acyclovir (antitherpetic drug)
Famciclovir (antitherpetic drug)
Valcyclovir (antitherpetic drug)
Interferon $\alpha$ -n3

Abbreviations: DES, diethylstilbestrol; SERMs, selective estrogen-response modulators.

following: *a*) smaller amounts of the test compound are required for study; *b*) housing and animal care expenses are less than those for other rodents; and *c*) faster breeding to generate multigenerational studies is possible. Reproductive tract development is similar in mice and humans. Therefore, the DES mouse model can be used to study developmental exposure to a wide range of compounds to which pregnant women may

be exposed including selective estrogen response modulators (SERMs).

### Animal Models That Will Accommodate SERMs

The activity of SERMs can be studied in animal models to assess compounds such as clomiphene or tamoxifen (43). Potential SERM activity of a compound may induce a response in the human that is not induced in

the animal model. This potential for false negative and false positive results can be reduced by using multiple animal models (mouse and rat) rather than relying on a single model in which SERM activity may be expressed. The danger is that all models may not be equally sensitive. Also, it is unclear whether a negative response is a result of decreased sensitivity. Although SERMs may be capable of inducing selective responses in specific animals, tissues, or conditions, as far as is known, all SERM activities have in common the binding to the ER. The use of receptor-binding assays is another approach to reliably screen for the effect of SERMs. These assays may include relative binding affinity analysis.

### What Are the Most Sensitive End Points?

Assays for the most sensitive end points must be able to detect the low-dose ranges that are well defined for DES. In addition, the full dose-response range must be determined for each compound and end point. The low-dose range for DES effect is approximately 0.02 µg/kg/day for the fetal mouse prostate (25) and 0.01 µg/kg/day for the neonatal mouse uterus (20,21,23). Overall, this low-dose range is defined as estrogenic activity delivered to cells in approximately the same low range of natural estrogen (e.g., free estradiol) at physiologic levels. A procedure and an approach for a predicted dose at this level have been described (26,44). It is not known if human fetal reproductive tract tissues are as sensitive to DES as are fetal mouse tissues. The nude mouse/human fetal tissue transplant system may provide data on this critical issue.

### Biomarkers

#### Molecular Markers

Patterning of the male and female genital tract has recently been shown to involve expression of *hox* and *wnt* genes (45–48). *hox* gene knockout (KO) studies have demonstrated profound disturbances in organogenesis and differentiation of the genital tract (46). DES has been shown to perturb the expression of several *hox* genes in the fetal Mullerian ducts when injected into pregnant mice (49). It will be useful to explore whether other exogenous estrogenic compounds share this activity. This new area of investigation requires further exploration to elucidate the role of these patterning genes in urogenital tract development in the mouse. Comparable studies in the human fetal reproductive tract are possible using the human fetal genital tract transplants to nude mice and would be desirable.

## High Sensitivity Biomarkers of DES Action

Lactoferrin is a protein that is regulated by estrogen in the female mouse reproductive tract. In the uterus of the adult mouse, lactoferrin transcripts are stimulated approximately 300-fold by estradiol or DES (50, 51). The expression of lactoferrin is induced by DES in the uterus prenatally, neonatally, and in adult stages (52), thus making it a particularly attractive biomarker of DES action in the mouse. Lactoferrin has also been localized to a subset of epithelial cells of the human endometrium and is responsive to estrogen (53). Comparable markers for the human uterus following exposure to other endocrine-disrupting chemicals should be sought.

## Engineered Assay Systems

Transgenic mice could be created using the lactoferrin promoter linked to green fluorescent protein. Estrogen action could be detected by external viewing through the body wall or via an intravaginal light detection system. In theory a single mouse could be used sequentially to test a series of separate compounds for their estrogenicity. This area of investigation is promising.

## What Can Be Learned Mechanistically for the DES Data?

A central question is which DES effects are dependent upon ER $\alpha$ -mediated or ER $\beta$ -mediated mechanisms. Studies with ER $\alpha$ -KO, ER $\beta$ -KO, and ER $\alpha$ /ER $\beta$  double KO mice will be required to settle these issues. Teratogenicity and/or carcinogenicity could be ER $\alpha$ -mediated either in the initiation or promotional phases. If tumor or birth defect initiation is an ER $\alpha$ -dependent event, the ER $\alpha$ -KO mouse will be useful in verifying that the initiating event involves estrogenic compounds acting through ER $\alpha$ . If tumor or birth defect initiation is an ER $\beta$ -dependent event, the ER $\beta$ -KO mouse will be useful in verifying that the initiating event involves estrogenic compounds acting through ER $\alpha$ . More sophisticated models could be created in which perinatal DES exposure can be achieved in an ER $\alpha$ -KO context with ER $\beta$  fully active to initiate lesions in the absence of ER $\alpha$ . At a later point it would be possible to selectively reestablish ER $\alpha$ -mediated estrogenic sensitivity by splicing out a stop cassette to reconstitute ER $\alpha$ . Through use of the CRE-lox system in transgenic mice, it should be possible to create such a model. By similar methods the role of ER $\beta$  in estrogen-induced teratogenicity and/or carcinogenicity should be explored. Studies on genetic and epigenetic changes associated with DES exposure will also be useful in understanding

the mechanism of action of DES and other endocrine-disrupting chemicals.

Collectively, the DES database in humans and rodents is likely to provide a highly relevant yardstick upon which to judge the potential estrogenic effects of compounds identified as environmental estrogens. The combination of known estrogenic potency with dose-response data for potential end points of concern in the mice and humans will be a useful tool for characterizing the effects of endocrine disrupters on human health at environmental exposure levels.

## REFERENCES AND NOTES

- Gardner WU. Estrogens in carcinogenesis. Arch Pathol 27:138-170 (1939).
- Geschickter CF. Diseases of the Breast. Philadelphia: J.B. Lippincott, 1943.
- Shimkin MB. Hormones and mammary cancer in mice. In: Mammary Tumors in Mice (Moulton FR, ed). Washington, DC: American Association for the Advancement of Science, 1945:85-122.
- Greene RR, Burrill MW, Ivy AC. The effects of estrogens on the antenatal sexual development of the rat. Am J Anat 67:305-345 (1939).
- Greene RR. Hormonal factors in sex inversion: the effects of sex hormones on embryonic sexual structures of the rat. Biol Symp 9:105-123 (1940).
- Karnaky KJ. Estrogenic tolerance of pregnant women. Am J Obstet Gynecol 53:312-316 (1947).
- Fraenkel L, Papanicolaou GN. Growth, desquamation and involution of the vaginal epithelium of fetuses and children with a consideration of the related hormonal effects. Am J Anat 62:427-451 (1938).
- Herbst AL, Ulfelder H, Poskanzer DC. Adenocarcinoma of the vagina: association of maternal stilbestrol therapy with tumor appearance in young women. N Engl J Med 284:878-881 (1971).
- Yonemura CY, Cunha GR, Sugimura Y, Mee SL. Temporal and spatial factors in diethylstilbestrol-induced squamous metaplasia in the developing human prostate. II: Persistent changes after removal of diethylstilbestrol. Acta Anat (Basel) 153:1-11 (1995).
- Taguchi O, Cunha GR, Robboy SJ. Experimental study of the effect of diethylstilbestrol on the development of the human female reproductive tract. Int J Biol Res Pregnancy 4:56-70 (1983).
- Taguchi O, Cunha GR, Lawrence WD, Robboy SJ. Timing and irreversibility of Mullerian duct inhibition in the embryonic reproductive tract of the human male. Dev Biol 106:394-398 (1984).
- Robboy SJ, Taguchi O, Cunha GR. Normal development of the human female reproductive tract and alterations resulting from experimental exposure to diethylstilbestrol. Hum Pathol 13:190-198 (1982).
- Sugimura Y, Cunha GR, Yonemura CU, Kawamura J. Temporal and spatial factors in diethylstilbestrol-induced squamous metaplasia in the developing human prostate. Hum Pathol 19:133-139 (1988).
- Brandenberger AW, Tee MK, Lee JY, Chao V, Jaffe RB. Tissue distribution of estrogen receptors  $\alpha$  (ER- $\alpha$ ) and  $\beta$  (ER- $\beta$ ) mRNA in the midgestational human fetus. J Clin Endocrinol Metab 82:3509-3512 (1997).
- Glatstein IZ, Yeh J. Ontogeny of the estrogen receptor in the human fetal uterus. J Clin Endocrinol Metab 80:958-964 (1995).
- Taguchi O, Cunha GR, Robboy SJ. Expression of nuclear estrogen-binding sites within developing human fetal vagina and urogenital sinus. Am J Anat 177:473-480 (1986).
- Kuiper GG, Enmark E, Pelto-Huikko M, Nilsson S, Gustafsson JA. Cloning of a novel receptor expressed in rat prostate and ovary. Proc Natl Acad Sci USA 93:5925-5930 (1996).
- Majumder PK, Kumar VL. Androgen receptor mRNA detection in the human foetal prostate. Int Urol Nephrol 29:633-635 (1997).
- Levine AC, Wang JP, Ren M, Eliashvili E, Russell DW, Kirschenbaum A. Immunohistochemical localization of steroid 5  $\alpha$ -reductase 2 in the human male fetal reproductive tract and adult prostate. J Clin Endocrinol Metab 81:384-389 (1996).
- McLachlan JA, Newbold RA, Shah HC, Hogan MD, Dixon RL. Reduced fertility in female mice exposed transplacentally to diethylstilbestrol (DES). Fertil Steril 38:364-371 (1982).
- McLachlan JA, Newbold RR, Bullock BC. Long-term effects on the female mouse genital tract associated with prenatal exposure to diethylstilbestrol. Cancer Res 40:3988-3999 (1980).
- Newbold R. Cellular and molecular effects of developmental exposure to diethylstilbestrol: implications for other environmental estrogens. Environ Health Perspect 103:83-87 (1995).
- Halling A, Forsberg JG. Acute and permanent growth effects in the mouse uterus after neonatal treatment with estrogens. Reprod Toxicol 7:137-153 (1993).
- vom Saal FS, Cooke PS, Buchanan DL, Palanza P, Thayer KA, Nagel SC, Parmigiani S, Welshons WV. A physiologically based approach to the study of bisphenol A and other estrogenic chemicals on the size of reproductive organs, daily sperm production, and behavior. Toxicol Ind Health 14:239-260 (1998).
- vom Saal FS, Timms BG, Montano MM, Palanza P, Thayer KA, Nagel SC, Dhar MD, Ganjam VK, Parmigiani S, Welshons WV. Prostate enlargement in mice due to fetal exposure to low doses of estradiol or diethylstilbestrol and opposite effects at high doses. Proc Natl Acad Sci USA 94:2056-2061 (1997).
- Nagel SC, vom Saal FS, Thayer KA, Dhar MG, Boechler M, Welshons WV. Relative binding affinity-serum modified access (RBA-SMA) assay predicts the relative *in vivo* bioactivity of the xenoestrogens bisphenol A and octylphenol. Environ Health Perspect 105:70-76 (1997).
- Smith OW, Smith GVS, Hurwitz D. Increased excretion of pregnanediol in pregnancy from diethylstilbestrol with special reference to the prevention of late pregnancy accidents. Obstet Gynecol 51:411-415 (1946).
- Golden RJ, Noller KL, Titus-Ernstoff L, Kaufman RH, Mittendorf R, Stillman R, Reese EA. Environmental endocrine modulators and human health: an assessment of the biological evidence. Crit Rev Toxicol 28:109-227 (1998).
- Kaufman RH, Adam E, Binder GL, Gerthoffer EA. Upper genital tract changes and pregnancy outcome in offspring exposed *in utero* to diethylstilbestrol. Am J Obstet Gynecol 154:330-332 (1990).
- Kaufman RH, Noller AE, Irwin JF, Gray M. Upper genital tract changes and infertility in diethylstilbestrol-exposed women. Am J Obstet Gynecol 154:1312-1318 (1986).
- Herbst A, Bern H. Developmental Effects of DES in Pregnancy. New York: Thieme Stratton, 1981.
- Herbst AL, Anderson D. Clear cell adenocarcinoma of the vagina and cervix secondary to intrauterine exposure to diethylstilbestrol. Semin Surg Oncol 6:343-346 (1990).
- Hatch EE, Palmer JR, Titus-Ernstoff L, Noller KL, Kaufman RH, Mittendorf R, Robboy SJ, Hyer M, Cowan CM, Adam E, Colton T, Hartzel P, Hoover RN. Cancer risk in women exposed to diethylstilbestrol *in utero*. JAMA 280:630-634 (1998).
- Jensen TK, Toppari J, Keiding N, Skakkebaek NE. Do environmental estrogens contribute to the decline in male reproductive health? Clin Chem 41:1896-1901 (1995).
- Bibbo M, Gill WB, Azizi F, Blough R, Fang VS, Rosenfield RL. Follow-up study of male and female offspring of DES-exposed mothers. Obstet Gynecol 49:1-8 (1977).
- Gill W, Schumacher G, Hubby M, Blough R. Male genital tract changes in humans following intrauterine exposure to diethylstilbestrol. In: Developmental Effects of Diethylstilbestrol (DES) in Pregnancy (Herbst A, Bern HA, eds). New York: Thieme Stratton Inc, 1981:103-119.
- Vingerhoets AJ, Assies J, Goodkin K, Van Heck GL, Bekker MH. Prenatal diethylstilbestrol exposure and self-reported immunorelated diseases. Eur J Obstet Gynecol Reprod Biol 77:205-209 (1998).
- Bekker MH, Heck GL, Vingerhoets AJ. Gender-identity, body-experience, sexuality, and the wish for having children in DES-daughters. Womens Health 24:65-82 (1996).
- Hornsby PP, Wilcox AJ, Weinberg CR, Herbst AL. Effects on the menstrual cycle of *in utero* exposure to diethylstilbestrol. Am J Obstet Gynecol 170:709-715 (1994).
- Marselos M, Tomatis L. Diethylstilbestrol. I: Pharmacology, toxicology and carcinogenicity in humans. Eur J Cancer 28A:1182-1189 (1992).
- Newbold R, Hanson RB, Jefferson WN, Bullock BC, Haseman J, McLachlan JA. Increased tumors but uncompromised fertility in the female descendants of mice exposed developmentally to diethylstilbestrol. Carcinogenesis (in press).
- Colborn T, vom Saal FS, Soto AM. Developmental effects of endocrine-disrupting chemicals in wildlife and humans. Environ Health Perspect 101:378-384 (1993).
- Newbold RR, Jefferson WN, Padilla-Burgos E, Bullock BC. Uterine carcinoma in mice treated neonatally with tamoxifen. Carcinogenesis 18:2293-2298 (1997).
- Nagel SC, vom Saal FS, Welshons WV. The effective free fraction of estradiol and xenoestrogens in human serum measured by whole cell uptake assays: physiology of delivery

- modifies estrogenic activity. *Proc Soc Exp Biol Med* 217:300–309 (1998).
45. Dolle P, Izpisua-Belmonte JC, Brown JM, Tickle C, Duboule D. HOX-4 genes and the morphogenesis of mammalian genitalia. *Genes Dev* 5:1767–1777 (1991).
  46. Warot X, Fromental-Ramain C, Fraulob V, Chambon R, Dolle P. Gene dosage-dependent effects of the Hoxa-13 and Hoxd-13 mutations on morphogenesis of the terminal parts of the digestive and urogenital tracts. *Development* 124:4781–4797 (1997).
  47. Pavlova A, Boutin E, Cunha GR, Sassoon D. Msx1 (Hox-7.1) in the mouse uterus: cellular interactions underlying regulation of expression. *Development* 120:335–346 (1994).
  48. Taylor HS, Vanden Heuvel GB, Igarashi P. A conserved Hox axis in the mouse and human female reproductive system: late establishment and persistent adult expression of the Hoxa cluster genes. *Biol Reprod* 57:1338–1345 (1997).
  49. Taylor KS, Block K, Kardana A, Igarashi P. *In utero* DES exposure alters Hox gene expression in the developing mouse Mullerian system. *J Society Gynecol Invest* 5 (suppl):39A (1998).
  50. Teng CT, Walker MP, Bhattacharyya SN, Klapper DG, DiAugustine RP, McLachlan JA. Purification and properties of an oestrogen-stimulated mouse uterine glycoprotein (approx. 70 kDa). *Biochem J* 240:413–422 (1986).
  51. Teng CT, Pentecost BT, Chen YH, Newbold RR, Eddy EM, McLachlan JA. Lactotransferrin gene expression in the mouse uterus and mammary gland. *Endocrinology* 124:992–999 (1989).
  52. Newbold RR, Teng CT, Beckman WC Jr, Jefferson WN, Hanson RB, Miller JV, McLachlan JA. Fluctuations of lactoferrin protein and messenger ribonucleic acid in the reproductive tract of the mouse during the estrous cycle. *Biol Reprod* 47:903–915 (1992).
  53. Walmer DK, Padin CJ, Wrona MA, Healy BE, Bentley RC, Tsao MS, Kohler MF, McLachlan JA, Gray KD. Malignant transformation of the human endometrium is associated with overexpression of lactoferrin messenger RNA and protein. *Cancer Res* 55:1168–1175 (1995).